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Phylogenetic analysis of *Theileria orientalis* in cattle bred in Fujian province, China

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ABSTRACT

A molecular epidemiological survey on *Theileria orientalis* was conducted in a cattle population of Fujian province in China. The screening polymerase chain reaction assay showed that 23 (45%) of 51 blood samples were positive for *T. orientalis*. DNA sequencing of the major piroplasm surface antigen-encoding gene indicated the presence of 3 different genotypes in the study area (types 1, 2, and 5) and identified *T. orientalis* types 1 and 2 as the major genotypes in the sample population. Because type 2 (Ikeda type in Japan) has been recognized as a relatively virulent genotype of *T. orientalis*, control and preventive measures to minimize the incidence of infectious diseases among cattle bred in China are necessary.

Keywords: *Theileria orientalis*; cattle; China; epidemiology; genotype

Theileria parasites are broadly classified into 2 groups. The first group consists of *Theileria parva* and *Theileria annulata* is known to cause malignant disease (Onuma *et al.*, 1997), whereas the second group consists of non-lymphoproliferative species (and classification and nomenclature systems have differed between authors) such as *Theileria orientalis* and *Theileria sergenti* (Kim *et al.*, 1998; Minami *et al.*, 1980). Although the second group has been described as having low pathogenicity, occasional outbreaks of infectious disease caused by the members of this group have been reported in many Asian cattle populations (Baek *et al.*, 1990). The clinical picture of *T. orientalis* infection may include anemia and related signs, and asymptomatic infections have been reported (Kawazu *et al.*, 1992).

Recently, the occurrence of different genotypes of *T. orientalis* has been widely reviewed, by using the gene encoding the major piroplasm surface protein (MPSP) of *T. orientalis* as an epidemiological molecular marker (Kakuda *et al.*, 1998; Khukhuu *et al.*, 2010). Previous classification criteria divided the parasite population into 4 genotypes, *i.e.*, I, C, B, and Thai (Kakuda *et al.*, 1998; Sarataphan *et al.*, 1999, 2003). Currently, however, the scheme proposed by Kim *et al.* (1998) has been increasingly adopted for phylogenetic analysis of *T. orientalis* isolates. According to this method, the *T. orientalis* population was initially classified into 6 genotypes (types 1-6) (Kim *et al.*, 1998). Subsequently, 2 more genotypes (types 7 and 8) were identified (Jeong *et al.*, 2010; Kim *et al.*, 1998; Ota *et al.*, 2009), and the recent analysis of the MPSP gene sequences of Vietnamese *T. orientalis* revealed 3 additional genotypes (types N1, N2 and N3) in the population (Kawazu *et al.*, 1999; Khukhuu *et al.*, 2010).

Most of these genotypes differ in antigenic structure; therefore, immunity against infection by a particular genotype may not provide protection against subsequent infection by other genotypes (Jeong *et al.*, 2010). The polymorphic nature of *T. orientalis* MPSP genes may hinder the development of a subunit vaccine, although immunization with recombinant MPSP has reduced the level of parasitemia (Onuma *et al.*, 1997). Therefore, regular, widespread epidemiological studies are necessary to identify the current distribution of different *T. orientalis* genotypes in a particular region or country, and to understand the phylogenetic associations between the different genotypes. Consequently, appropriate preventive and control strategies can be formulated against *T. orientalis* infection. Recently, we conducted molecular epidemiological studies on *T. orientalis* found in the circulating blood of domestic animals bred in various

countries (Khukhuu *et al.*, 2010; Ota *et al.*, 2009; Yokoyama *et al.*, 2011). In addition, a recent epidemiological survey indicated the presence of 6 *T. orientalis* genotypes (types 1-5 and 7) in China (Liu *et al.*, 2011). In this study, we investigated a cattle population bred in China and analyzed the *MPSP* gene to identify the molecular epidemiology of *T. orientalis* in the study area, Fujian.

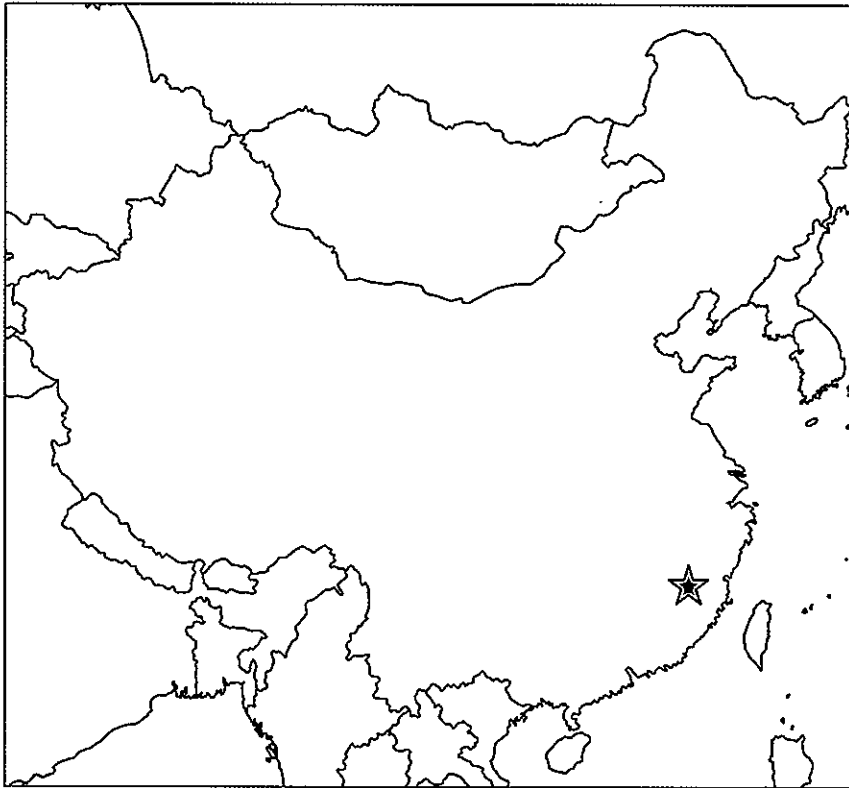


Figure 1: Location of Fujian province in China(★). Blood samples were collected from 51 randomly selected cattle bred in the province.

We randomly collected 51 blood samples from a cattle population bred in Fujian province (Fig. 1), on October 20, 2009, and extracted DNA with a commercial kit (Qiagen Blood Mini Kit, Hilden, Germany). The DNA samples were then screened for the presence of *T. orientalis* by a previously described diagnostic polymerase chain reaction (PCR) assay (Ota *et al.*, 2009). In the PCR assay, a set of forward and reverse primers was used for amplification of the *T. orientalis*-specific *MPSP* gene fragment (Ota *et al.*, 2009). Briefly, 25 μ L of the reaction mixture consisted of 1 μ L DNA template, 2.5 μ L $\times 10$ reaction buffer (Applied Biosystems, Branchburg, NJ, USA), 2.5 μ L 2 mM dNTPs (Applied Biosystems), 1 μ L 10 μ M primers each, 0.1 μ L *Taq* DNA polymerase (Applied Biosystems), and 16.9 μ L double distilled water. The reaction was amplified under optimized conditions as described previously (Ota *et al.*, 2009). Identification of a 776 bp PCR product on gel electrophoresis indicated the presence of *T. orientalis* in the sample population (Fig. 2), and of 51 DNA samples, 23 samples (45%) were positive in the PCR assay (Table 1). Although this positive percentage was higher than that of previous investigations in China (34%) (Liu *et al.*, 2011), limited sample numbers in this study might account for the observed difference.

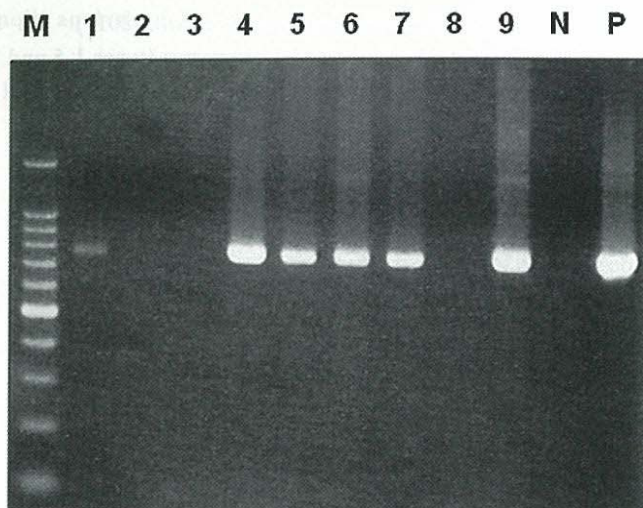


Figure 2: PCR detection of *T. orientalis* genes. M, 100 bp DNA marker; lanes 1–9, field DNA samples extracted from bovine blood; N, negative control; P, positive control.

Table 1. PCR detection and genotyping of *T. orientalis* detected from cattle bred in Fujian province, China.

Total number of collected samples	Number of positive samples(%)	Isolated genotypes (Number) ¹⁾
51	23(45.0)	1(7), 2(7), 5(4)

¹⁾ Refers to Figure 3.

Eighteen of the positively amplified PCR products were successfully gel extracted (Alquick Gel Extraction Kit, Qiagen), ligated to a PCR 2.1 plasmid vector (PCR 2.1-TOPO, Invitrogen, Carlsbad, CA, USA), cloned, and then cultured as previously described (Ota *et al.*, 2009). Plasmids were extracted from the positive cultures to obtain the inserted fragment, and the nucleotide sequences of the inserted *MPSP* gene fragments were determined by a genetic analyzer (ABI PRISM 3100 genetic analyzer, Applied Biosystems). The sequences were then analyzed, according to previously documented methods (Ota *et al.*, 2009). Briefly, after multiple alignments with the ClustalW online software program, a phylogenetic tree was constructed, based on a neighbor-joining branching pattern (Perrière *et al.*, 1996). Additionally, a bootstrap test was employed to determine the reliability of the branching pattern of the phylogenetic tree (Felsenstein, 1985).

The phylogenetic analysis identified 3 different genotypes, *i.e.*, types 1, 2 and 5 (Fig. 3), and identified *T. orientalis* types 1 and 2 as the major genotypes in the study area. Type 1 (Chitose type in Japan) has already been reported in China and other countries such as Russia, Japan and Korea (Gubbels *et al.*, 2000), and recently, we indicated the presence of type 1 in Mongolia (Altengerel *et al.*, 2011), Thailand (Altengerel *et al.*, 2011) and Vietnam (Khukhuu *et al.*, 2010). Type 2 (Ikeda type in Japan) has been isolated in China, Japan and Korea (Kakuda *et al.*, 1998; Kim *et al.*, 1998; Ota *et al.*, 2009; Yokoyama *et al.*, 2011). Type 5 was initially reported in Japan, Korea, and very recently in China (Jeong *et al.*, 2010; Kakuda *et al.*, 1998; Kim *et al.*, 1998; Liu *et al.*, 2011; Yokoyama *et al.*, 2011). In addition, type 5 has recently been isolated in Mongolia (Altengerel *et al.*, 2011), Thailand (Altengerel *et al.*, 2011) and Vietnam (Khukhuu *et al.*, 2010).



1

The results of our present study clearly indicate the presence of 3 different *T. orientalis* genotypes in the sample population, but a large-scale analysis of bovine and/or tick populations is required to generalize the findings of this study. Compared with the other *T. orientalis* genotypes, type 2 (Ikeda type) has been described as a virulent genotype in Japan (Kamau *et al.*, 2011). Because the circulating blood of the bovine population of Fujian province was found to harbor type 2 as one of the major genotypes, the cattle industry in the study area may be susceptible to economical losses due to *T. orientalis* infection. Therefore, an integrated control and preventive strategy must be developed to minimize disease incidence among the cattle bred in Fujian province, China.

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References

- Altangerel, K., Battsetseg, B., Battur, B., Sivakumar, T., Batnagnai, E., Javkhlan, G., Tuvshintulga, B., Batsukh, Z., Igarashi, I., Matsumoto, K., Inokuma, H. and Yokoyama, N. 2011. The first epidemiological survey of *Theileria orientalis* infection in Mongolian cattle. *Vet. Parasitol.* [In press].
- Altangerel, K., Sivakumar, T., Inpankaew, T., Jittapalpong, S., Terkawi, M.A., Ueno, A., Xuan, X., Igarashi, I. and Yokoyama, N. 2011. Molecular prevalence of different genotypes of *Theileria orientalis* detected from cattle and water buffaloes in Thailand. *J. Parasitol.* [In press].
- Baek, B.K., Kim, B.S. and Lee, H.I. 1990. Fine structures of *Theileria sergenti* in Korean native cattle. *Korean J. Vet. Res.* 30: 465-471.
- Felsenstein, J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. *Evolution.* 39: 783-791.
- Gubbels, M.J., Hong, Y., van der Weide, M., Qi, B., Nijman, I.J., Guangyuan, L. and Jongejan, F. 2000. Molecular characterisation of the *Theileria buffeli/orientalis* group. *Int. J. Parasitol.* 30: 943-952.
- Jeong, W., Yoon, S.H., An, D.J., Cho, S.H., Lee, K.K. and Kim, J.Y. 2010. A molecular phylogeny of the benign *Theileria* parasites based on major piroplasm surface protein (MPSP) gene sequences. *Parasitology.* 137: 241-249.
- Kakuda, T., Kubota, S., Sugimoto, C., Baek, B.K., Yin, H. and Onuma, M. 1998. Analysis of immunodominant piroplasm surface protein genes of benign *Theileria* parasites distributed in China and Korea by allele-specific polymerase chain reaction. *J. Vet. Med. Sci.* 60: 237-239.
- Kamau, J., de Vos, A.J., Playford, M., Salim, B., Kinyanjui, P. and Sugimoto, C. 2011. Emergence of new types of *Theileria orientalis* in Australian cattle and possible cause of theileriosis outbreaks. *Parasit. Vectors.* 4: 22.
- Kawazu, S., Sugimoto, C., Kamio, T. and Fujisaki, K. 1992. Antigenic differences between Japanese *Theileria sergenti* and other benign *Theileria* species of cattle from Australia (*T. buffeli*) and Britain (*T. orientalis*). *Parasitol. Res.* 78: 130-135.
- Kawazu, S., Kamio, T., Kakuda, T., Terada, Y., Sugimoto, C. and Fujisaki, K. 1999. Phylogenetic relationships of the benign *Theileria* species in cattle and Asian buffalo based on the major piroplasm surface protein (p33/34) gene sequences. *Int. J. Parasitol.* 29: 613-618.
- Khukhuu, A., Lan, D.T., Long, P.T., Ueno, A., Li, Y., Luo, Y., Macedo, A.C., Matsumoto, K., Inokuma, H.,

- Kawazu, S.I., Igarashi, I., Xuan, X. and Yokoyama, N. 2010. Molecular Epidemiological Survey of *Theileria orientalis* in Thua Thien Hue Province, Vietnam. J. Vet. Med. Sci. [In press].
- Kim, S.J., Tsuji, M., Kubota, S., Wei, Q., Lee, J.M., Ishihara, C. and Onuma, M. 1998. Sequence analysis of the major piroplasm surface protein gene of benign bovine *Theileria* parasites in east Asia. Int. J. Parasitol. 28: 1219-1227.
- Liu, A.H., Guan, G.Q., Liu, J.L., Liu, Z.J., Leblanc, N., Li, Y.Q., Gao, J.L., Ma, M.L., Niu, Q.L., Ren, Q.Y., Bai, Q., Yin, H. and Luo, J.X. 2011. Polymorphism analysis of Chinese *Theileria sergenti* using allele-specific polymerase chain reaction of the major piroplasm surface protein gene. J. Parasitol. 97: 116-121.
- Minami, T., Fujinaga, T., Furuya, K. and Ishihara, T. 1980. Clinico-hematologic and serological comparisons of Japanese and Russian strains of *Theileria sergenti*. Natl. Inst. Anim. Health. Q. (Tokyo). 20: 44-52.
- Onuma, M., Kubota, S., Kakuda, T., Sako, Y., Asada, M., Kabeya, H. and Sugimoto, C. 1997. Control of *Theileria sergenti* infection by vaccination. Trop. Anim. Health. Prod. 29: 119S-123S.
- Ota, N., Mizuno, D., Kuboki, N., Igarashi, I., Nakamura, Y., Yamashina, H., Hanzaike, T., Fujii, K., Onoe, S., Hata, H., Kondo, S., Matsui, S., Koga, M., Matsumoto, K., Inokuma, H. and Yokoyama, N. 2009. Epidemiological survey of *Theileria orientalis* infection in grazing cattle in the eastern part of Hokkaido, Japan. J. Vet. Med. Sci. 71: 937-944.
- Perrière, G. and Gouy, M. 1996. WWW-query: an on-line retrieval system for biological sequence banks. Biochimie. 78: 364-369.
- Sarataphan, N., Nilwarangkoon, S., Tananyutthawongese, C., Kakuda, T., Onuma, M. and Chansiri, K. 1999. Genetic diversity of major piroplasm surface protein genes and their allelic variants of *Theileria* parasites in Thai cattle. J. Vet. Med. Sci. 61: 991-994.
- Sarataphan, N., Kakuda, T., Chansiri, K. and Onuma, M. 2003. Survey of benign *Theileria* parasites of cattle and buffaloes in Thailand using allele-specific polymerase chain reaction of major piroplasm surface protein gene. J. Vet. Med. Sci. 65: 133-135.
- Yokoyama, N., Ueno, A., Mizuno, D., Kuboki, N., Khukhuu, A., Igarashi, I., Miyahara, T., Shiraishi, T., Kudo, R., Oshiro, M., Zakimi, S., Sugimoto, C., Matsumoto, K. and Inokuma, H. 2011. Genotypic diversity of *Theileria orientalis* detected from cattle grazing in Kumamoto and Okinawa prefectures of Japan. J. Vet. Med. Sci. 73: 305-312.